

## Facile Synthesis of Dioscin and Its Analogues

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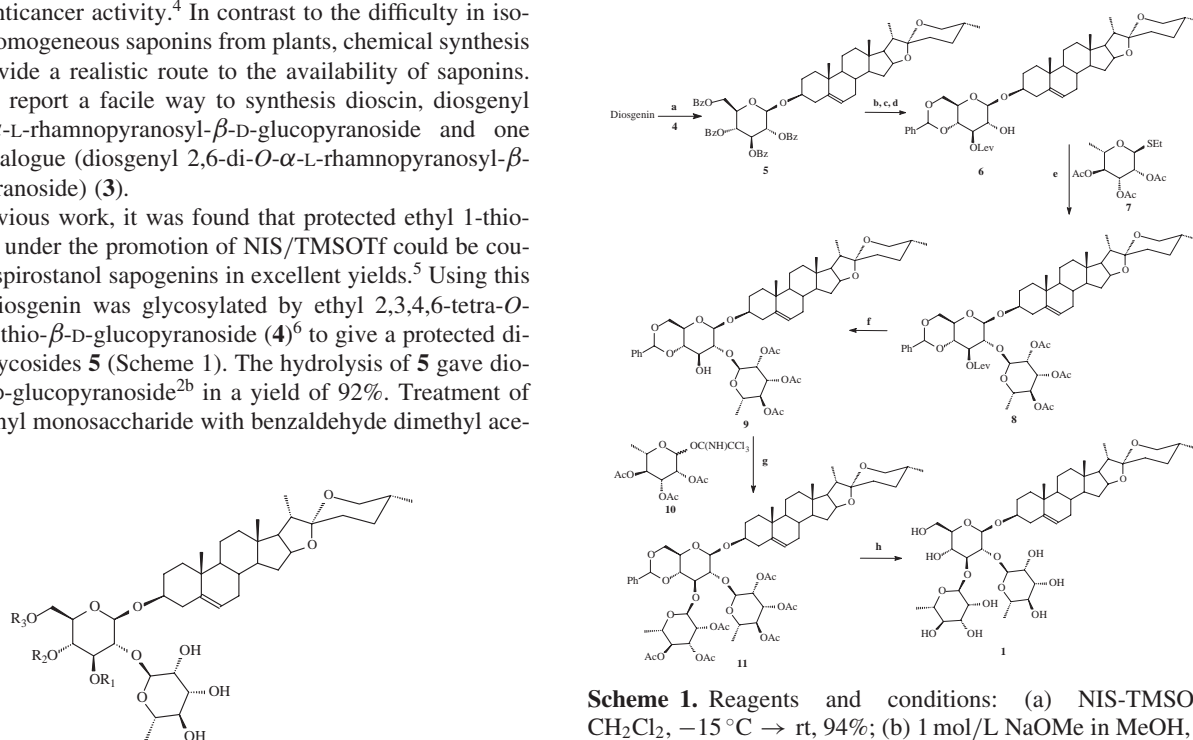
Three representative spirostanol saponins that have typical structure of the sugar moiety, diosgenyl 2,3-di-*O*- $\alpha$ -L-rhamnopyranosyl- $\beta$ -D-glucopyranoside, diosgenyl 2,4-di-*O*- $\alpha$ -L-rhamnopyranosyl- $\beta$ -D-glucopyranoside (dioscin), and diosgenyl 2,6-di-*O*- $\alpha$ -L-rhamnopyranosyl- $\beta$ -D-glucopyranoside, were synthesized in a facile way. An approach to selectively mask the C<sub>3</sub>-hydroxyl of diosgenyl 4,6-*O*-benzylidene  $\beta$ -D-glucopyranoside was described. A procedure using cerium(IV) ammonium nitrate for selective removal of *tert*-butyldimethylsilyl group while retaining levulinyl group is afforded.

Saponins, a structurally and biologically diverse class of glycosides, are major component in traditional Chinese medicines.<sup>1</sup> The structure diversity of saponins lies mainly in their sugar moieties.<sup>1</sup> Many studies have disclosed that some biological functions of saponins can be ascribed to the sapogenin and some to the carbohydrate residue,<sup>2</sup> for example diosgenyl glucosides. Dioscin (**2**) displays cardiovascular and antitumor activities.<sup>3</sup> Moreover, recent study had found that diosgenyl 2,3-di-*O*- $\alpha$ -L-rhamnopyranosyl- $\beta$ -D-glucopyranoside (**1**) also showed stronger anticancer activity.<sup>4</sup> In contrast to the difficulty in isolation of homogeneous saponins from plants, chemical synthesis would provide a realistic route to the availability of saponins. Herein we report a facile way to synthesis dioscin, diosgenyl 2,3-di-*O*- $\alpha$ -L-rhamnopyranosyl- $\beta$ -D-glucopyranoside and one of their analogue (diosgenyl 2,6-di-*O*- $\alpha$ -L-rhamnopyranosyl- $\beta$ -D-glucopyranoside) (**3**).

In previous work, it was found that protected ethyl 1-thioglycosides under the promotion of NIS/TMSOTf could be coupled with spirostanol sapogenins in excellent yields.<sup>5</sup> Using this method, diosgenin was glycosylated by ethyl 2,3,4,6-tetra-*O*-benzoyl-1-thio- $\beta$ -D-glucopyranoside (**4**)<sup>6</sup> to give a protected diosgenyl glycosides **5** (Scheme 1). The hydrolysis of **5** gave diosgenyl  $\beta$ -D-glucopyranoside<sup>2b</sup> in a yield of 92%. Treatment of the diosgenyl monosaccharide with benzaldehyde dimethyl ace-

tal and a catalytic amount of *p*-toluenesulfonic monohydrate in DMF gave diosgenyl 4,6-*O*-benzylidene  $\beta$ -D-glucopyranoside. The Lev group was introduced by reaction of diosgenyl 4,6-*O*-benzylidene  $\beta$ -D-glucopyranoside with levulinic acid and DCC in the presence of a catalytic amount of DMAP. The desired compound 3-*O*-Lev **6** was afforded in a yield of 73%, although it was quite difficult to selectively mask one of the hydroxyl groups of the 2,3-diol of D-glucopyranose, especially when it was in the  $\beta$ -form.<sup>7</sup> NIS/TMSOTf mediate coupling of ethyl 2,3,4-tri-*O*-acetyl-1-thio- $\alpha$ -L-rhamnopyranose (**7**)<sup>8</sup> with **6**, gave **8** in a yield of 87%. It was anticipated that treatment with hydrazine acetate in CH<sub>2</sub>Cl<sub>2</sub>/MeOH would cleave the Lev without the effect to other acetyl groups. Indeed, compound **8** was converted into **9** within 2 h in a yield of 90%. Sugar receptor **9** was glycosylated by 2,3,4-tri-*O*-acetyl-L-rhamnopyranosyl trichloroimidate (**10**)<sup>9</sup> to give the protected diosgenyl trisaccharides **11** in a yield of 92%. Treatment of **11** with 80% HOAc at 70 °C for 5 h, then removal of all of the acetyl groups with MeONa furnished **1** in a yield of 85%.

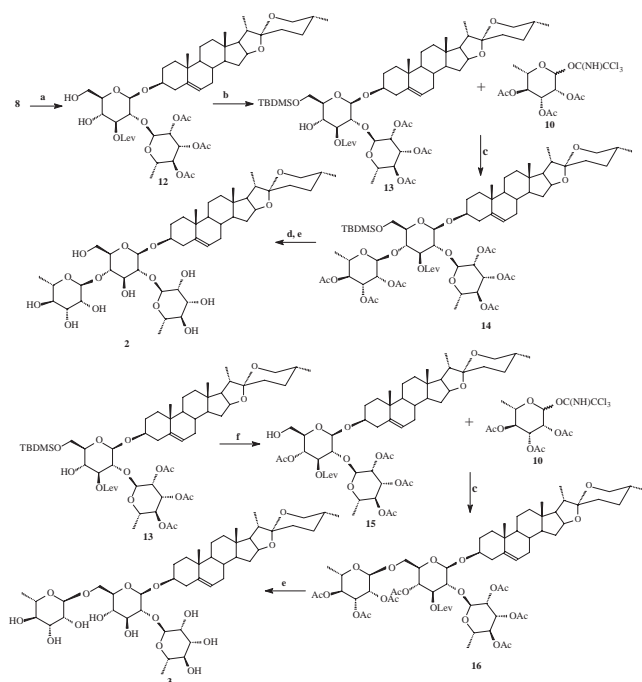
With compound **8** in hand, dioscin (**2**) and diosgenyl 2,6-di-



- $R_1 = \alpha$ -L-rhamnopyranosyl,  $R_2 = R_3 = H$ ;
- $R_2 = \alpha$ -L-rhamnopyranosyl,  $R_1 = R_3 = H$ ;
- $R_3 = \alpha$ -L-rhamnopyranosyl,  $R_1 = R_2 = H$ ;

**Figure 1.** The structure of dioscin and its analogues.

**Scheme 1.** Reagents and conditions: (a) NIS-TMSOTf, CH<sub>2</sub>Cl<sub>2</sub>, -15 °C  $\rightarrow$  rt, 94%; (b) 1 mol/L NaOMe in MeOH, reflux, 92%; (c) PhCH(OMe)<sub>2</sub>, DMF, *p*-TsOH, (d) levulinic acid, DCC, DMAP, CH<sub>2</sub>Cl<sub>2</sub>, rt, 73%; (e) NIS-TMSOTf, CH<sub>2</sub>Cl<sub>2</sub>, -30 °C  $\rightarrow$  rt, 87%; (f) H<sub>2</sub>NNH<sub>2</sub>-HOAc, CH<sub>2</sub>Cl<sub>2</sub>-MeOH, rt, 90%; (g) BF<sub>3</sub>·Et<sub>2</sub>O, CH<sub>2</sub>Cl<sub>2</sub>, -40 °C  $\rightarrow$  rt, 92%; (h) 80% HOAc, 5 h, 70 °C and then MeONa, CH<sub>2</sub>Cl<sub>2</sub>-MeOH, rt, 85%.



**Scheme 2.** Reagents and conditions: (a) 80% HOAc, 3 h, 80 °C, 92%; (b) imidazole, DMAP, TBDMSiCl, DMF, rt, 93%; (c) BF<sub>3</sub>·Et<sub>2</sub>O, CH<sub>2</sub>Cl<sub>2</sub>, -40 °C → rt, 58% for **14**, 80% for **17**; (d) CAN, CH<sub>2</sub>Cl<sub>2</sub>-MeOH, rt, 95%; (e) 1 mol/L NaOMe in MeOH, rt, 81% for **2**, 89% for **3**; (f) Ac<sub>2</sub>O, pyridine, rt, 5 h and then 80% HOAc, 1 h, 70 °C, 78%.

*O*- $\alpha$ -L-rhamnopyranosyl- $\beta$ -D-glucopyranoside (**3**) were synthesized as shown in Scheme 2. Treatment of **8** with 80% HOAc at 80 °C for 3 h provided compound **12** in a yield of 92%. In the presence of imidazole, DMAP and TBDMSiCl, the C<sub>6</sub>'-OH of **12** was selectively protected and gave compound **13** in a yield of 93%. Though 2.0 equiv. TBDMSiCl of **12** had been used, the product of silylation on C<sub>4</sub>'-OH was not found. Using compound **13** as sugar acceptor to couple with sugar donor **10**, protected diosgenyl trisaccharide **14** was afforded in a yield of 58%. The low yield of this glycosylation could be caused by the steric hindrance of the C<sub>4</sub>'-OH in **13**. During the process of cleavage TBDMS from **14** in the presence of TBAF in THF, the Lev group was also removed off. The reason was probably contributed to the instability of Lev to F<sup>-</sup> caused by TBAF.<sup>10</sup> Alternatively, treatment with CAN<sup>11</sup> in MeOH/CH<sub>2</sub>Cl<sub>2</sub>, TBDMS of **14** was cleanly cleaved within 1 h and no loss of the Lev group was observed. The hydrolysis of the intermediate using MeONa/MeOH, dioscin (**2**) was produced in 76% yield of two steps. Acetylation of **13** and then treatment with 80% HOAc at 80 °C for 1 h, furnished **15** in a yield of 78% overall two steps. Glycosylation with sugar donor **10** provided compound **16**. Deprotection of acyl groups with MeONa, afforded target saponin **3** in a yield of 89%.

In conclusion, regioselective protection C<sub>3</sub>-hydroxyl of diosgenyl 4,6-*O*-benzylidene  $\beta$ -D-glucopyranoside by levulinyl group enables compound **8** to be produced in a better yield. Taken **8** as the synthon, three sugar acceptors **9**, **13**, and **15** were gotten respectively in high yields. Therefore, dioscin and its analogues were facily prepared in satisfactory overall yields.

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- The spectral data of **1**: <sup>1</sup>H NMR (500 MHz, pyridine-*d*<sub>5</sub>): 5.86 (brs, 1H, H-1'''), 5.79 (brs, 1H, H-1''), 5.29 (d, *J* = 4.5 Hz, 1H, H-6), 4.90–4.83 (m, 3H), 4.80–4.76 (m, 2H), 4.55–4.20 (m, 4H), 4.34–4.27 (m, 3H), 4.16 (t, *J* = 8.5 Hz, 1H), 4.07–4.03 (m, 2H), 3.91–3.87 (m, 1H), 3.79–3.77 (m, 1H), 3.58–3.46 (m, 2H), 2.74–2.64 (m, 2H), 2.08–1.99 (m, 2H), 1.93 (m, 1H), 1.73 (d, *J* = 6.0 Hz, 3H), 1.64 (d, *J* = 5.5 Hz, 3H), 1.13 (d, *J* = 7.0 Hz, 3H), 1.02 (s, 3H), 0.81 (s, 3H), 0.67 (d, *J* = 5.0 Hz, 3H). <sup>13</sup>C NMR (75 MHz, pyridine-*d*<sub>5</sub>): 140.73, 121.81, 109.23, 103.84, 102.61, 99.86, 87.41, 81.09, 78.40, 78.08, 77.75, 73.80, 73.57, 72.81, 72.62, 72.51(2C), 70.58, 69.87, 66.84, 62.87, 62.24, 56.61, 50.24, 41.95, 40.44, 39.83, 38.64, 37.46, 37.10, 32.29, 32.20, 31.81, 31.66, 30.58, 30.04, 29.25, 21.08, 19.37, 18.66, 18.41, 17.31, 16.32, 15.02. ESI-MS: 891.6 (M + Na)<sup>+</sup>. HRMS (FAB-MS): calcd for C<sub>45</sub>H<sub>72</sub>O<sub>16</sub>Na (M + Na)<sup>+</sup>: 891.4718, found: 891.4725.
- The spectral data of **2** as reported before: C.-C. Zou, S.-J. Hou, Y. Shi, P.-S. Lei, and X.-T. Liang, *Carbohydr. Res.*, **338**, 721 (2003).
- The spectral data of **3**: <sup>1</sup>H NMR (500 MHz, pyridine-*d*<sub>5</sub>): 6.35 (brs, 1H, H-1'''), 5.48 (brs, 1H, H-1''), 5.30 (d, *J* = 4.5 Hz, 1H, H-6), 4.98–4.95 (m, 2H), 4.81 (d, *J* = 3.5 Hz, 1H), 4.63–4.61 (dd, *J* = 3.5, 9.0 Hz, 1H), 4.59–4.48 (m, 4H), 4.37–4.32 (m, 2H), 4.24–4.16 (m, 2H), 4.06 (m, 1H), 3.97–3.92 (m, 2H), 3.59–3.48 (m, 3H), 2.80–2.70 (m, 2H), 2.22 (m, 1H), 1.77 (d, *J* = 6.5 Hz, 3H), 1.63 (d, *J* = 6.0 Hz, 3H), 1.13 (d, *J* = 7.0 Hz, 3H), 1.00 (s, 3H), 0.80 (s, 3H), 0.68 (d, *J* = 5.0 Hz, 3H). <sup>13</sup>C NMR (75 MHz, pyridine-*d*<sub>5</sub>): 140.84, 121.63, 109.21, 102.41, 102.17, 100.83, 81.07, 79.53, 78.69, 78.01, 76.55, 74.06, 73.98, 72.79, 72.68, 72.51, 72.22, 71.80, 69.77, 69.51, 67.91, 66.81, 62.84, 56.52, 50.12, 41.91, 40.39, 39.79, 39.16, 37.45, 37.04, 32.17, 31.77, 31.61, 30.55, 30.33, 29.95, 29.57, 29.21, 21.02, 19.35, 18.64, 17.30, 16.29, 15.02. ESI-MS: 891.6 (M + Na)<sup>+</sup>, HRMS (FAB-MS): calcd for C<sub>45</sub>H<sub>72</sub>O<sub>16</sub>Na (M + Na)<sup>+</sup>: 891.4718, found: 891.4697.